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Introduction

Recently, development of oligonucleotide therapeutics is accelerated in pharmaceutical industry, and the needs for their ultrasensitive quantitation methods are greatly increased. Conventional approaches, such as hybridization, LC-MS/MS, PCR are used in the analysis. Hybridization is commonly used, yet sensitivity is insufficient. Sensitivity of LC-MS/MS is improved by instrumental advancement, but still not enough. PCR and qRT-PCR are also known as sensitive detection methods; however, the high variability of measured values is a problem. To solve these problems, we aimed to develop an ultrasensitive hybridization assay method without nucleic acid amplification procedure.

PALSAR

PALSAR (Probe Alternation Link Self-Assembly Reaction) is a signal amplification technology, based on self-assembly of a DNA probe pair. PALSAR does not need DNA/RNA polymerases and ligases. In this study, we assayed by Luminex-PALSAR method, a combination of Luminex technology enabling multiplex detection (xMAP) and PALSAR.

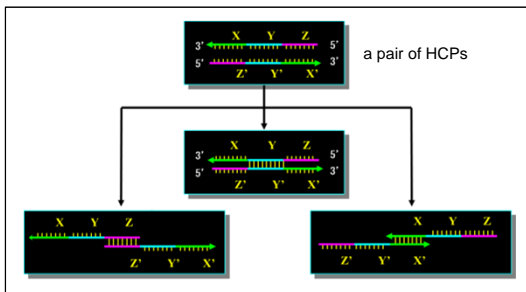


Figure 1. Principle of PALSAR

Honeycomb probes (HCPs) are DNA probes and have 3 regions, X, Y, Z for the one probe, and X', Y', Z' for the other probe. The regions X and X', Y and Y', and Z and Z' are complementary to each other, therefore three kinds of dimers will be formed.

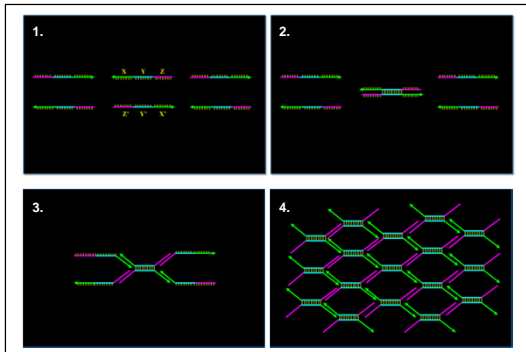
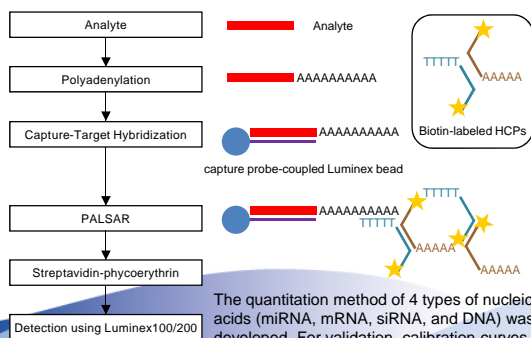


Figure 2. Illustrations of PALSAR

(1) A pair of HCPs is used for PALSAR. (2) One of the 3 regions in the probe is hybridized with the complementary region of the other probe. (3) The rest of the regions in the probes are hybridized. (4) A self-assembly is formed by PALSAR.

Workflow



The quantitation method of 4 types of nucleic acids (miRNA, mRNA, siRNA, and DNA) was developed. For validation, calibration curves, intra-day reproducibility, and inter-day reproducibility were studied.

Results

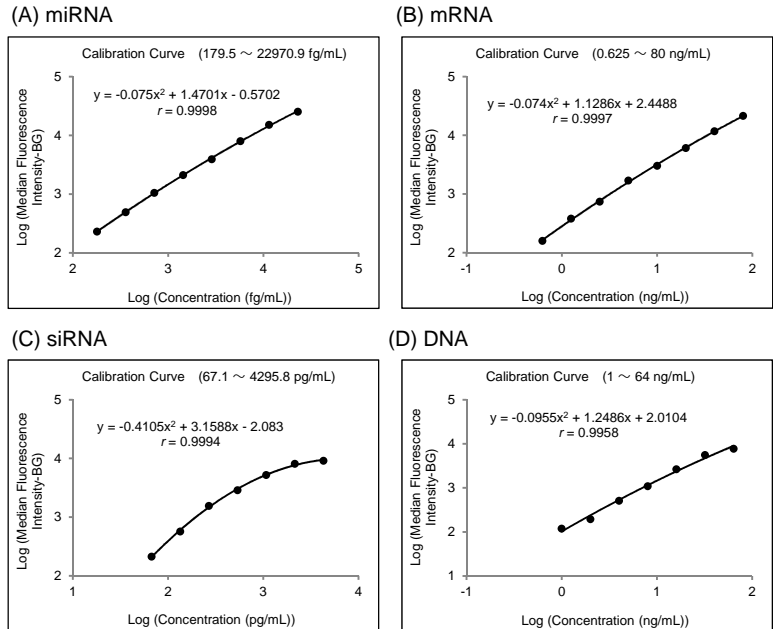


Figure 3. Representative calibration curves

(A) 179.5 to 22970.9 fg/mL of miRNA (22 bases) (custom-synthesized by Takara Bio, HPLC grade) was prepared with TE (10 mM Tris-HCl, 1 mM EDTA, pH 8.0). (B) 0.625 to 80 ng/mL of mRNA (1,380 bases) (purchased from NIPPON GENE) was prepared with nuclease-free water. (C) 67.1 to 4295.8 pg/mL of siRNA (19 bp with dTdT 3' overhang) (custom-synthesized by Nihon Gene Research Laboratories, HPLC grade) was prepared with nuclease-free water. (D) 1 to 64 ng/mL of phosphorothioated DNA (20 bp) (purchased from Gene Design) was prepared with nuclease-free water.

Table 1. Assessment of calibration curve of miRNA

| Nominal concentration (fg/mL) | RE (%) | | | | | | <i>r</i> | | | | | |
|-------------------------------|--------|-------|-------|-------|-------|-------|----------|--------|--------|--------|--------|--------|
| | Day 1 | Day 2 | Day 3 | Day 4 | Day 5 | Day 6 | Day 1 | Day 2 | Day 3 | Day 4 | Day 5 | Day 6 |
| 179.5 | 3.9 | 4.8 | -4.0 | 0.4 | 3.1 | -1.5 | 0.9997 | 0.9995 | 0.9991 | 0.9998 | 0.9991 | 0.9997 |
| 358.9 | -5.1 | -5.0 | 5.2 | -0.9 | -0.2 | 4.3 | | | | | | |
| 717.8 | -2.1 | -5.0 | 6.7 | 1.7 | -2.0 | 0.1 | | | | | | |
| 1435.7 | 1.8 | 5.2 | -3.7 | 0.4 | -8.8 | -6.1 | | | | | | |
| 2871.4 | 3.2 | -4.2 | -11.6 | -4.8 | -1.6 | 1.0 | | | | | | |
| 5742.7 | -3.0 | 5.7 | 7.8 | 1.9 | 13.7 | -0.4 | | | | | | |
| 11485.4 | 5.1 | 3.8 | 4.9 | 5.4 | 7.5 | 7.8 | | | | | | |
| 22970.9 | -3.1 | -5.0 | -2.8 | -3.1 | -9.3 | -4.7 | | | | | | |

A series of standard solutions for calibration curve was prepared as described in Figure 3. Assay of this study was performed for 6 days.

Table 2. Intra-day and inter-day reproducibility of miRNA

| n | Nominal concentration (fg/mL) | | | | | n=18 | Nominal concentration (fg/mL) | | | | |
|--------|--------------------------------|-------|--------|---------|---------|--|--------------------------------|-------|--------|---------|---------|
| | 179.5 | 358.9 | 2871.4 | 11485.4 | 22970.9 | | 179.5 | 358.9 | 2871.4 | 11485.4 | 22970.9 |
| | Observed concentration (fg/mL) | | | | | | Observed concentration (fg/mL) | | | | |
| 1 | 209.2 | 379.6 | 2736.3 | 11656.4 | 22062.4 | Mean | 175.3 | 334.0 | 2677.8 | 11581.9 | 21833.5 |
| 2 | 191.0 | 364.1 | 2845.3 | 13156.2 | 22195.7 | SD | 20.7 | 26.7 | 214.2 | 743.0 | 539.6 |
| 3 | 196.6 | 323.4 | 2722.4 | 11493.5 | 22279.0 | RE (%) | -2.3 | -6.9 | -6.7 | 0.8 | -5.0 |
| Mean | 198.9 | 355.7 | 2768.0 | 12102.0 | 22179.0 | CV (%) | 11.8 | 8.0 | 8.0 | 6.4 | 2.5 |
| SD | 9.3 | 29.0 | 67.3 | 916.6 | 109.3 | TE (%) | 14.1 | 14.9 | 14.7 | 7.2 | 7.5 |
| RE (%) | 10.8 | -0.9 | -3.6 | 5.4 | -3.4 | 179.5 to 22970.9 fg/mL of miRNA was prepared for quality control samples. Assay of this study was performed in triplicate. For inter-day reproducibility assessment, assay was performed for 6 days. | | | | | |
| CV (%) | 4.7 | 8.2 | 2.4 | 7.6 | 0.5 | | | | | | |
| TE (%) | 15.5 | 9.1 | 6.0 | 13.0 | 3.9 | | | | | | |

Conclusions

Luminex-PALSAR method, which has been developed and evaluated in this study, is able to quantitate various types of nucleic acids with ultra-sensitivity. Calibration ranges of nucleic acids are verified as follows: 179.5 to 22970.9 fg/mL miRNA; 0.625 to 80 ng/mL mRNA; 67.1 to 4295.8 pg/mL siRNA; 1 to 64 ng/mL DNA. This method can be expected to be a powerful technique for ultrasensitive quantitation of oligonucleotide therapeutics.

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We have no financial relationship to disclose for our presentation contents.